

MICROARRAY TECHNOLOGIES: BENCH TO BEDSIDE

DNA microarray technology has matured to the point where some applications are deemed reliable enough for use in patient care. At the same time, microarrays are evolving to help expand the understanding of transcriptome complexity: single nucleotide polymorphisms, copy number variation, CpG methylations, microRNAs—so many genetic and epigenetic variations and a slew of microarrays to investigate each one. Furthermore, newer DNA sequencing technologies now threaten to do to microarray technology what automobiles did to the horse and buggy. For now, however, the two approaches appear to coexist happily, and microarrays remain a perfectly reasonable way to get around the genome.

By Emma Hitt

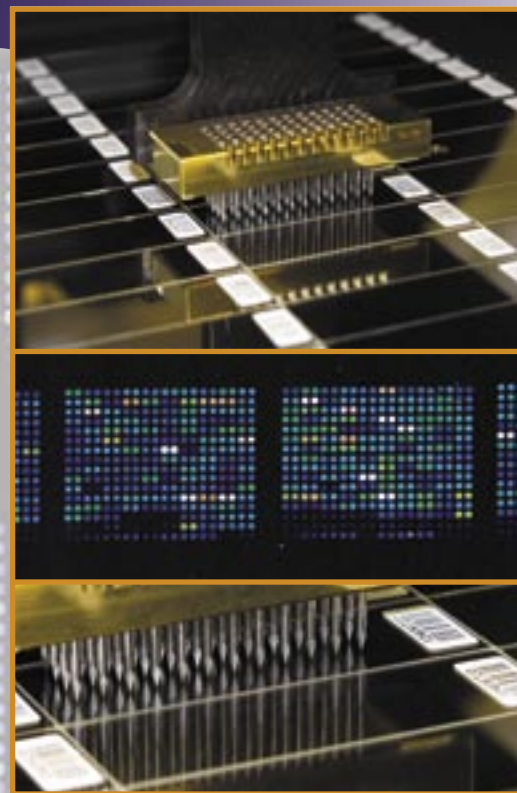
The first report on microarray technology appeared in *Science* barely more than a decade ago, and since its inception, microarray technology has been burdened with issues of reproducibility and standardization. Recently, however, the technology has received a vote of confidence in the form of US Food and Drug Administration approval, laying the groundwork for its broader application in a clinical setting. The newly approved test, the MammaPrint assay, was developed by **Agendia** in the Netherlands and measures the expression of 70 genes that predict the likelihood of breast cancer metastasis. Several such tests have already received the “CE mark” for clinical use in Europe, and a handful of tests that analyze DNA polymorphisms (as opposed to gene expression) have been available clinically in the United States since 2004, when **Roche’s** AmpliChip CYP450 assay became the first to receive approval.

As is the case for most biological paradigms, the entire transcriptome within a given cell has turned out to be much more complicated than ever imagined. As a result, tools to study epigenetic variations and other regulatory mechanisms within the cell have proliferated. Microarrays can now measure changes in gene copy number (copy number variation, CNV), changes in the expression levels of microRNAs (miRNAs), contributions from gene splice variants, and changes in methylation patterns. “There are really a plethora of new assay types, and each of these is complementary to one another and informs from a different perspective,” notes Kevin Meldrum, director of genomics marketing for **Agilent**.

DNA Sequencing Technologies

Perhaps one of the most exciting new developments in the microarray field is one that—paradoxically—may render the technology obsolete. DNA sequencing—the remarkably robust kind that has allowed a pioneer like James Watson to obtain his personal genome sequence on a DVD in about two months—is becoming the five-lane superhighway of genomic analysis. A 454 Life Sciences (now a part of Roche Diagnostics) sequencer was used to complete Watson’s analysis, but several other companies now have their own sequencing approaches including **Illumina’s** Solexa technology, **Applied Biosystems’** SOLiD System, **Helicos BioSciences’** True Single Molecule Sequencing (tSMS), and others.

“An interesting paradigm shift is taking place,” says Steve Lombardi, chief operating officer and executive vice president with Helicos BioSciences. “You can do virtually any genetic analysis with sequencing technology—ranging from quantification of DNA, RNA, or cDNA, to measuring copy number variation and methylation,” he says. “Microarrays, which require probes with known sequences, have limitations that simply do not exist with sequencing technology.” [continued >](#)



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Microarray Technologies

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According to Lombardi, True Single Molecule Sequencing has the advantage of a simple sample preparation process that does not require amplification, and the image processing software and informatics will be open source allowing users to innovate.

A popular application of sequencing technology is to replace classic microarray chromatin immunoprecipitation (ChIP)-chip analyses. Microarrays to measure protein binding to DNA surfaced about five years ago and involve the placement of immunoprecipitated DNA fragments as probes on a DNA microarray. There is a wide range of products from Agilent, **Affymetrix**, Illumina, **Aviva Systems Biology**, **Nimblegen Systems**, and others for this purpose. With sequencing technology, the whole process becomes scaled up and digitized. First, binding proteins are cross-linked in place to DNA, the chromatin is isolated and fragmented, a protein-specific antibody is added, and the protein-DNA complexes are purified. The binding proteins are then chemically removed, and the remaining DNA is isolated. The DNA strands are then size selected, a process that isolates short sequences of, say, 25 bases—millions and millions of them. Finally, binding sequences are identified by virtue of the fact that their sequences are enriched several-fold relative to other sequences.

“What this massively parallel approach to sequencing allows us to do is to generate about a hundred-fold more data for about 1 percent of the cost of current technology,” notes Omead Ostadan vice president of marketing with Illumina. Some proponents suggest that the sequencing approach will ultimately replace microarrays. “Microarrays are most popular for performing gene expression and genotyping applications, and in some cases, for sequencing applications such as ChIP-chip analysis. Over time some of these applications may migrate over to a sequencing platform because of the added power and quality you get from sequencing. However, we see these two approaches as complementary, offering researchers the broadest solutions for studies,” says Todd Dickinson, director of product marketing at Illumina.

It may not be prudent to throw out the array chips just yet, however. “Sequencing technology is young, and issues regarding cost and ease of use still need to be worked out,” notes Steven Bodovitz, an industry analyst with **BioPerspectives**. “Nonetheless, sequencing is certainly more comprehensive than microarray technology, and getting a sequence without knowing what you are looking for ahead of time is a big advantage,” he says. “It will be interesting to see how the two approaches play out—the technologies are likely to both complement and compete with one another.”

Still, microarrays remain ideally suited for experiments that require analysis of large numbers of samples against known sequences—and will likely remain the predominant approach in diagnostic applications. “Not all companies are jumping on board with the sequencing approach,” says Bodovitz.

Optimizing Arrays

Of late, developments in gene expression arrays have focused on optimizing coverage of the entire human genome, which includes

well over 30,000 genes, processed into hundreds of thousands of different variations. Newer arrays are aiming to capture that variability. Affymetrix’s GeneChip Human Gene 1.0 ST Array released this year measures the overall expression of all transcripts derived from a gene, in contrast to traditional arrays that measure only the 3’ end of a gene. “More than 60 percent of human genes undergo alternative splicing, resulting in multiple transcript variants with potentially distinct functions,” says Yan Zhang, associate director of product marketing with Affymetrix. Each of 28,869 genes is represented on the array by approximately 26 probes per gene directed at the full length of the gene. “We estimate that the accuracy is greater compared with the previous generation’s 3’ approach, and the format is smaller, which reduces cost without reducing content,” Zhang says.

Illumina’s whole-genome expression arrays employ the BeadArray technology, which uses 50-mer oligos attached to 3 μm beads, allowing six or eight samples to be profiled per BeadChip simultaneously. This multisample approach provides increased throughput and decreased cost per sample. Also allowing for higher throughput and lower costs, Nimblegen’s Human Whole Genome Expression Microarrays include 60-mer oligo probes for all human genes, eight per gene, including 47,633 different targets, and Agilent has a whole human genome chip containing 60-mer oligonucleotide probes arranged as microarrays in the format of 1 x 244K, 2 x 105K, 4 x 44K, or 8 x 15K on individual glass slides.

Microarray Manufacture

Arrays can be loosely classified into ready-made versions and custom “home-brew” forms that contain DNA spotted onto slides. Custom microarrays are fabricated largely by pin-based direct deposition printers.

UK-based company **Arrayjet** recently launched its benchtop Sprint Inkjet Microarrayer. The new machine can handle 20 microarray slides and allows printing from two microtiter plates (96- or 384-well) simultaneously.

“Due to their contact with the surface, pin printers can have problems with speed, quality, and reproducibility,” notes Howard Manning, Arrayjet founder and technical director. “With the ink jet approach, the Jetspyder enables the inkjet print head to draw in multiple samples before arraying spots are printed on the slide without the print head stopping and without making contact,” says Manning. Another advantage of inkjet printing is that spotting pins may damage some of the newer substrates used for arrays, such as membrane-coated substrates and hydrogels.

The processes utilized to manufacture arrays are also being enhanced. “Updates to the Agilent SurePrint technology were implemented last year,” Meldrum said, “enabling us to manufacture arrays with up to 244,000 unique features. We have also improved the efficiency of the nucleic acid synthesis process, significantly reducing depurination side reactions.” **Nanogen** NanoChip arrays use a proprietary technology consisting of individually controllable microelectrodes that, when positively charged, can attract negatively charged DNA and RNA molecules. “The use of electronics to drive the concentration reaction mitigates the variability arising from spotting small volumes onto a surface,” says Nanogen’s Elaine Weidenhammer. “Consequently, the Nanogen system is subject to less variability than some other formats,” she claims.

SNPs and CNV

A recent publication from The ENCYCLOPEDIA OF DNA ELEMENTS (ENCODE), an international research consortium organized by the National Human Genome Research Institute (NHGRI) indicates that the DNA-makes-RNA-makes-protein paradigm is too simplistic and that 99 percent of the genome is not, in fact, junk. **continued** ▶

Gene transcripts appear to overlap, and genes, regulatory elements, and other types of DNA sequences interact as a network in a nonlinear fashion. New technologies are now evolving to define this complex variability.

This year, Affymetrix introduced the Genome-Wide Human SNP Array 6.0, which includes about 1.8 million markers that can be used to detect genetic variation in whole-genome association studies. Specifically, the newest SNP Array 6.0 contains 906,600 single nucleotide polymorphisms (SNPs) and about 946,000 nonpolymorphic probes that “will aim to detect CNVs in both known and unknown regions,” says Jessica Tonani, Genotyping Specialist with Affymetrix. “The array tiles known CNVs very heavily, with an average of about 61 probes per known CNV region. For the remainder of the genome we have tiled probes at a resolution of about a probe every 700 bases.”

“Our overall hope is to identify the part of the genome that has previously been considered a ‘genomic wasteland,’ or junk DNA, but may in fact be involved in regulating gene expression,” she says, “although we are reaching a point where there is more to be gained by studying more samples as opposed to putting more SNPs on an array.”

Newest to Illumina’s Infinium portfolio is the Human 1M BeadChip, which provides expanded genomic coverage for whole-genome association studies. This chip adds several hundred thousand individually selected SNPs in genes and functional areas as well as CNV content not available on any other platform, according to Illumina. Tag SNPs represent a group of SNPs that tend to be inherited together. “By targeting tag SNPs, we can vastly increase the power of the study by looking at the genotype of one SNP, which effectively represents the genotype of many,” Dickinson says. “The power of this array is increased dramatically because of this coverage and has reduced costs for researchers conducting genome-wide association studies,” he adds. “For example, in 2000, genotyping a single SNP

cost \$1 or more. Today, the cost is less than \$0.001 per SNP.”

Regulatory miRNA

MiRNAs have recently been shown to play an important role in regulating gene expression. Over 500 human miRNAs have already been identified and validated, and are thought to regulate about one-third of all human genes. Several companies now make kits for labeling and detection of miRNAs utilizing array platforms. Agilent recently introduced its miRNA assay which uses a direct labeling protocol on as little as 100 ng of total RNA. “Because miRNAs are typically 25–30 nucleotides in length and have high sequence homology, utilization of conventional probe design methods results in significant cross-hybridization of RNAs within the sample,” notes Agilent’s Meldrum, “but we have developed novel probe design algorithms that enable us to achieve extremely high specificity for each unique sequence.”

Kreatech Biotechnology in the Netherlands recently introduced its miRacULS II Kit labeling technology, which uses a nonenzymatic labeling reaction independent of fragment size. The kit enables isolation and labeling of miRNAs from samples of less than 105 cells or 5 mg tissue as well as samples with a low miRNA expression level.

Ready for Prime Time?

Efforts are ongoing to standardize data input and reporting of microarray experimental data. **Minimal Information about Microarray Experiment (MAIME)** standards for microarray experiments have been put in place to define the minimal amount of information needed to interpret and reproduce a microarray experiment.

The first phase of the **MicroArray Quality Control (MAQC)** project, with participants from academia, government, and industry, recently concluded that results mostly were reproducible among labs; the second phase of the MAQC is under way and will try to establish the applicability of microarray data to clinical settings, with results expected in about a year. “When moving something from the bench to medical practice, there is an absolute requirement that the consistency of the performance of the test system can be guaranteed in different people’s hands,” says Uwe Scherf, with the FDA’s Center for Devices and Radiological Health. “The MAQC studies were one of the first indications that microarrays could be suitable for medical evaluation.”

The External RNA Controls Consortium (ERCC), established in 2003, is developing a set of external RNA spike-in controls for use in all microarray platforms and quantitative polymerase chain reaction (PCR) assays. If successful, this will allow a reliable comparison of expression profiling among more than 10 microarray and PCR platforms.

According to Affymetrix, at least 20 diagnostic tests are being developed for use on its platform—mostly expression-based tests evaluating tumors—and they are partnering with several companies, including Roche, **Almac Diagnostics**, **Veridex**, **BioMerieux**, and **Pathwork Diagnostics**, for the development of molecular diagnostic tests that use its platform. VeraCode, a new technology introduced by Illumina earlier this year, is available for researchers conducting low- and midplex biomarker validation studies. Using digital holographic codes embedded in cylindrical glass beads, VeraCode allows for efficient and cost-effective screening of hundreds to thousands of samples against panels of one to 384 biomarkers.

“It will be important to develop tools that can be utilized to analyze data sets for different applications and start to merge that information together,” Agilent’s Meldrum says. “Within the next few years the field is going to go from one-dimensional [e.g., gene expression profiling] to a multidimensional view, offering a richer picture of what’s really happening in the cell.”

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Helicos BioSciences Corporation

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MicroArray Quality Control (MAQC)

www.fda.gov/nctr/science/centers/toxicoinformatics/maqc

Minimal Information about Microarray Experiment (MIAME)

www.mged.org/Workgroups/MIAME/miame.html

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Microarray Data Analysis

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Correction

Life Science Technologies Feature:

"Cell Signaling: Phosphorylation, Kinases, and Kinase Inhibitors" (6 April 2007, p. 125). The trademark for the homogeneous time-resolved fluorescence (HTRF) technology was incorrectly attributed to Millipore Corporation. The HTRF trademark is held by Cisbio International of Gif-sur-Yvette, France.